

## Biosynthesis of Quinones of *Streptocarpus dunnii* Cell Cultures

Kenichiro Inoue, Shinichi Ueda, Hidekazu Nayeshiro, and Hiroyuki Inouye\*

Faculty of Pharmaceutical Sciences, Kyoto University, Sakyo-ku, Kyoto 606, Japan

Three unusually prenylated naphthoquinones (**1**), (**5**), and (**6**) of *Streptocarpus dunnii* are found to be biosynthesised *via* (**2**), (**8**), (**9**), and the Claisen type rearrangement product (**10**) of the latter, while two anthraquinones (**3**) and (**4**) of the same plant are shown to be formed in a prenylation mode different from that of the structurally related anthraquinones of Rubiaceae plants.

We have reported the isolation of six dunnione (**1**) type naphthoquinones and two anthraquinones from *Streptocarpus dunnii* Mast. (Gesneriaceae) and its cell cultures.<sup>1</sup> These naphthoquinones feature an unusual binding mode of the prenyl group to the quinone nucleus, which probably stems from 4-(2-carboxyphenyl)-4-oxobutanoic acid (**2**) already known as a precursor of one of the anthraquinones (**3**).<sup>2</sup> These anthraquinones are structurally congeneric to those of Rubiaceae plants.<sup>3</sup>

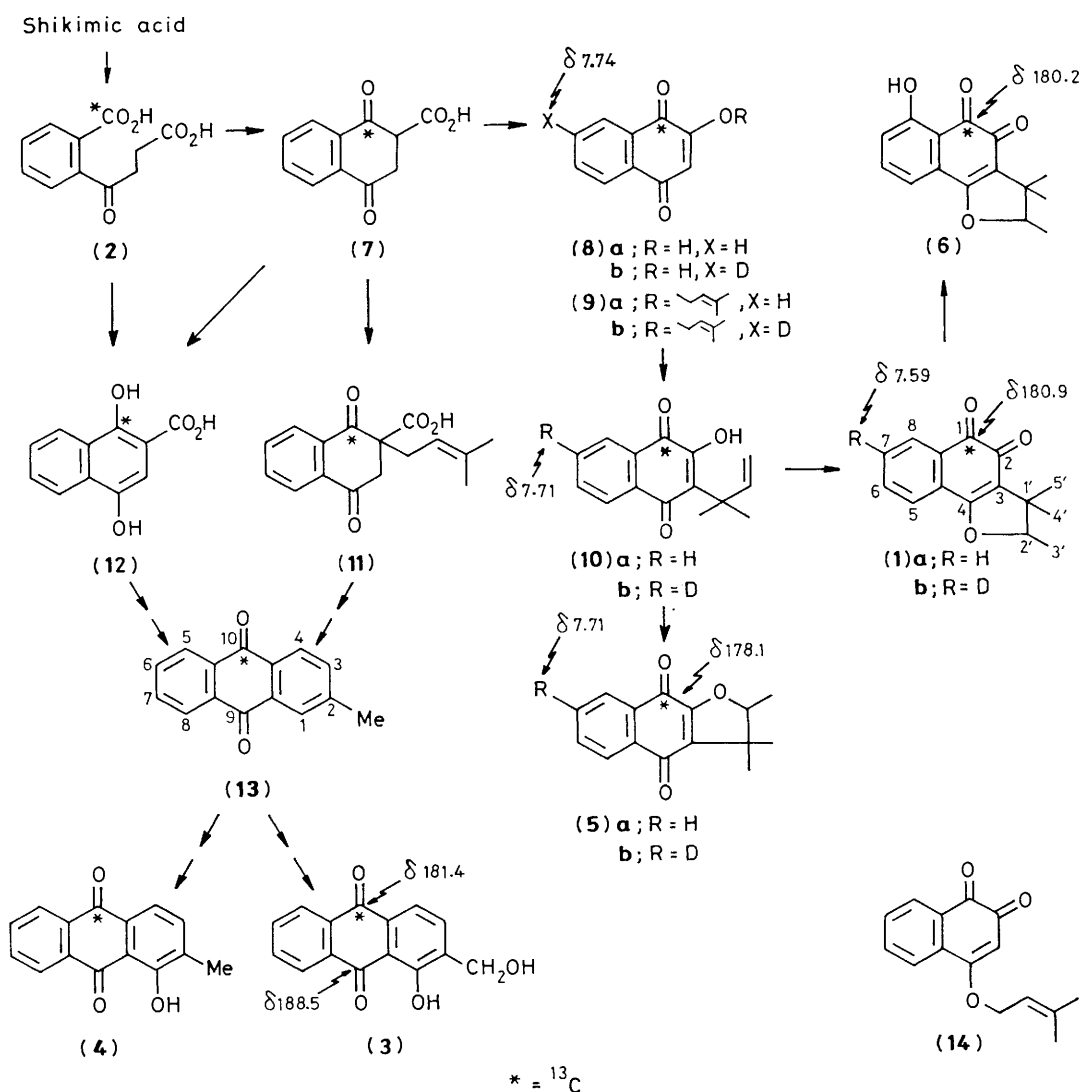
We therefore studied the biosynthesis of these quinones by feeding experiments with <sup>13</sup>C and <sup>2</sup>H labelled precursors. *S. dunnii* cell suspension cultures coexisting with half differentiated plantlets were used as plant material. They were incubated in Linsmaier-Skoog medium supplemented with 10<sup>-5</sup> M indoleacetic acid and 10<sup>-6</sup> M kinetin at 25 °C in the dark for one month after subculturing.

[2-carboxy-<sup>13</sup>C]-4-(2-Carboxyphenyl)-4-oxobutanoic acid (**2**)<sup>3</sup> (50 mg, 86.5 atom %) was fed to the cultures (5 × 170 ml medium) and incubated under the same conditions for an additional 2 weeks. The combined benzene extracts of the cells and plantlets as well as the medium were chromatographed on silica gel to give dunnione (**1a**) (5.2 mg), 1-hydroxy-2-hydroxymethylanthraquinone (**3**) (30.3 mg), 1-hydroxy-2-methylanthraquinone (**4**) (3.1 mg), α-dunnione (**5a**) (10.0 mg), and 8-hydroxydunnione (**6**) (6.3 mg). The <sup>13</sup>C n.m.r. spectra of these quinones indicated that the label was introduced into the 1-position for naphthoquinones and the 10-position for anthraquinones. The enrichment factors of <sup>13</sup>C in the quinones (**1a**), (**3**), (**4**), (**5a**), and (**6**) calculated on the basis of the mass spectra were 11.6, 20.7, 9.5, 19.5, and 7.7 atom % excess, respectively. Therefore, it was verified that

all these quinones were formed *via* (**2**). The location of the introduced label in the quinones led to the following conclusions. (i) The naphthoquinones are biosynthesised *via* 2-carboxy-2,3-dihydro-1,4-naphthoquinone (**7**) and lawsone (**8a**);<sup>4</sup> the latter is prenylated at the C-2 position to form the ether (**9a**) followed by a Claisen-type rearrangement to afford the naphthoquinone (**10a**), which is further converted into (**1a**), (**5a**), and (**6**). (ii) The anthraquinones are not formed *via* the C-3 prenylation as in Rubiaceae plants;<sup>3</sup> they are formed *via* (**7**) and the prenylation product (**11**),<sup>5</sup> as for the naphthoquinone congeners of *Catalpa ovata*, or *via* the naphthoic acid (**12**),<sup>6</sup> as for the menaquinones of *Mycobacterium phlei*.

The intermediacy of lawsone (**8**) in the biosynthesis of naphthoquinones was proved in the following way. Synthesised [7-<sup>2</sup>H]-lawsone (**8b**)† (25 mg, 94.0 atom %) was administered to the cultures (5 × 170 ml) under the conditions described above, and after two weeks incubation, the usual constituents (**1b**) (0.7 mg), (**3**) (21.3 mg), (**4**) (2.0 mg), (**5b**) (12.7 mg), and (**6**) (6.5 mg) along with tectoquinone (**13**) (1.0 mg) were isolated. The mass spectra of these compounds indicated that the label was incorporated into naphthoquinones (**1b**), (**5b**), and (**6**) with enrichment factors of 17.5, 24.1, and 0.8 atom % excess, respectively, and no label was introduced into the anthraquinones (**3**), (**4**), and (**13**). The <sup>2</sup>H n.m.r. spectrum of (**5b**) showed that the label was on C-7, establishing the intermediacy of (**8**). The unusual production of (**13**), which may be caused by a feedback control from the

† The synthesis of [7-<sup>2</sup>H]-lawsone (**8b**) starting from 4-(4-chlorophenyl)butanoic acid will be reported elsewhere.



Scheme 1

addition of (8), supported the intermediacy of (13) in the biosynthesis of (3) and (4).

The intermediacy of lawsone 2-prenyl ether (9) was then examined. The [7-<sup>2</sup>H<sub>1</sub>]-ether (9b) (50 mg) derived from [7-<sup>2</sup>H<sub>1</sub>]-lawsone (8b) was fed to the cultures (5 × 170 ml) and the quinones were isolated as before. In this experiment, besides the constituents (1b) (5.5 mg), (3) (16.5 mg), (4) (2.0 mg), (5b) (12.2 mg), (6) (3.7 mg), and (13) (2.3 mg), the intermediate (10b) (31 mg) was also obtained. Its structure was established by an alternative synthesis. The enrichment factors of <sup>2</sup>H in the quinones (1b), (5b), and (6) were 58.2, 65.3, and 2.2 atom % excess, respectively, whereas the value for (10b) was 82.6 atom % excess. This unusually high value and the high yield of (10b) can be explained by the overflow production caused by the excessive addition of the precursor (9b). The label in the quinones (1b), (5b), and (10b) was shown to be at C-7 from their <sup>2</sup>H n.m.r. spectra. The above results unequivocally demonstrate that naphthoquinones of the cell cultures of *S. dunnii* are biosynthesised *via* (8), (9), and (10). The production of the intermediate (10) on addition of (9) excludes the possibility that the quinones are formed *via* direct prenylation of (8) in an inverted mode, or *via* the Claisen-type rearrangement of 1,2-naphthoquinone 4-prenyl ether (14), which might be formed from (8).

On the basis of these results, the biosynthetic pathway shown in Scheme 1 is proposed for quinones of cell cultures of *S. dunnii*. The extremely low enrichment factors for 8-hydroxydunnione (6) compared to those of the other quinones in both <sup>2</sup>H feeding experiments can be attributed to the NIH shift<sup>7</sup> during the hydroxylation of (1).

Received, 15th June 1982; Com. 683

## References

- 1 K. Inoue, S. Ueda, H. Nayeshiro, and H. Inouye, *Chem. Pharm. Bull.*, 1982, **30**, 2265.
- 2 J. Stöckigt, U. Srocka, and M. H. Zenk, *Phytochemistry*, 1973, **12**, 2389.
- 3 K. Inoue, Y. Shiobara, H. Nayeshiro, H. Inouye, G. Wilson, and M. H. Zenk, *J. Chem. Soc., Chem. Commun.*, 1979, 957, and references cited therein.
- 4 K. Inoue, Y. Shiobara, and H. Inouye, *Chem. Pharm. Bull.*, 1977, **25**, 1462.
- 5 K. Inoue, S. Ueda, Y. Shiobara, I. Kimura, and H. Inouye, *J. Chem. Soc., Perkin Trans. 1*, 1981, 1246.
- 6 R. Meganathan and R. Bentley, *J. Bacteriol.*, 1979, **140**, 92, and references cited therein.
- 7 J. W. Daly, D. M. Jerina, and B. Witkop, *Experientia*, 1972, **28**, 1129.